EZassay

Nucleic Acid Extraction and Purification Kit

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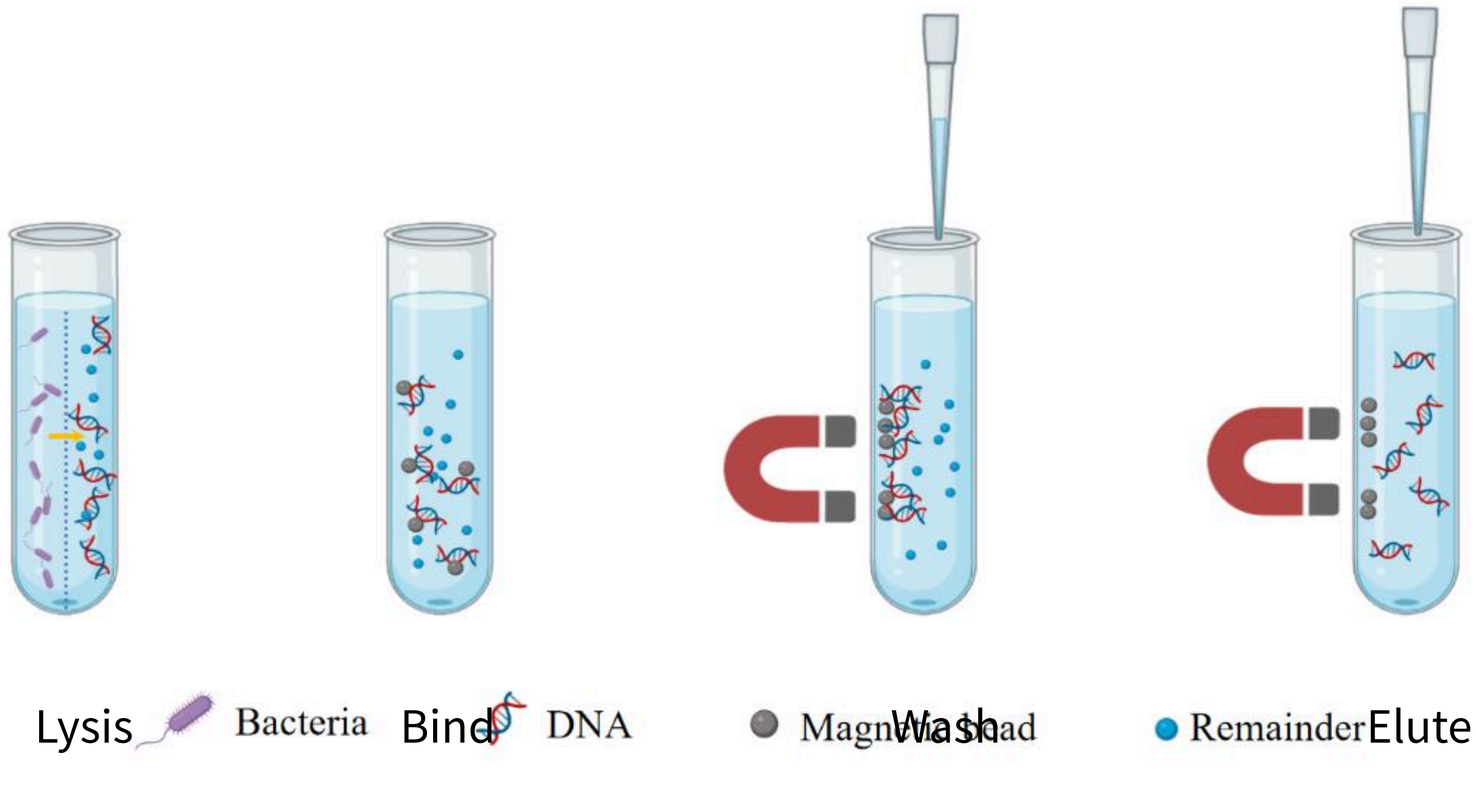
Catalog Code: MB-WG-050
MB-WG-200

CONTENTS

Brief introduction

The uniquely embedded magnetic beads have a strong affinity for nucleic acids under certain conditions, and when the conditions change, the magnetic beads release adsorbed nucleic acids, which can achieve the purpose of rapid isolation and purification of nucleic acids.



Application

This kit is intended for the extraction, enrichment, and purification of nucleic acids (DNA and RNA).

Materials supplied

Component	Cat. NO.： MB-WG-050 Package size=50 tests	Cat. NO.： MB-WG-200 Package size=200 tests	Store
Magnetic bead suspension	1ml	1ml*4	2~8℃
Lysis buffer*	15ml	60ml	Room temperature
Wash buffer A	16ml	16ml*4	Room temperature
Wash buffer B	20ml	20ml*4	Room temperature
Elution buffer	5ml	20ml	Room temperature

*Check the Lysis Solution for salt precipitation before each use. Re-dissolve any precipitate by warming the solution to 37°C, then mix well and cool to 25°C before use.

Required materials but not supplied

i. 100% isopropanol, molecular biology grade.
ii. 100% ethanol, molecular biology grade.
iii. Vortex.
iv. Centrifuge, 1.5 mL centrifuge tubes.
v. Pipettes and pipette tips.
vi. Magnetic particle processing rack.

Procedure

Ensure that ethanol was added to Wash Buffer A & Wash Buffer B before the first use.

Component	Wash buffer A	Wash buffer B
Concentrated buffer	16ml	20ml
100% Ethanol	64ml	60ml
Total	80ml	80ml

1. Take out the magnetic bead suspension from 2-8°C and equilibrate to room temperature. Mix and resuspend on a vortex to a visibly homogeneous suspension and ensure that no settling occurs prior use.
2. Transfer 200µl of sample to a 1.5ml centrifuge tube. Add 300µL of lysis buffer. Mix well by vortex. Briefly spin down the tube to collect droplets.
3. Add 20µl of magnetic bead suspension and 350µl of isopropanol. Incubate for 10 minutes in mixer. Briefly spin down the tube to collect droplets.
4. Place the centrifuge tube on the magnetic stand for 3 minutes, or until the beads have formed a tight pellet. Without removing the tube from the magnetic rack, remove and discard the supernatant carefully by using a pipette.
5. Add 700µl of wash buffer A. Mix well by using vortex for 1 minute. Briefly spin down the tube to collect droplets.
6. Place the centrifuge tube on the magnetic stand for 3 minutes, or until the beads have formed a tight pellet. Without removing the tube from the magnetic rack, remove and discard the supernatant carefully by using a pipette

7. Repeat steps 5 and 6;
8. Add 700µl of washing buffer B. Mix well by using vortex for 1 minute. Briefly spin down the tube to collect droplets.
9. Place the centrifuge tube on the magnetic stand for 3 minutes, or until the beads have formed a tight pellet. Without removing the tube from the magnetic rack, remove and discard the supernatant carefully by using a pipette.
10. Repeat steps 8 and 9;
11. Without removing the microcentrifuge tube from the magnetic rack, open the lid for 10 minutes at room temperature. Allow residual ethanol to volatilize.
(Note: avoid complete drying of the beads.)
12. Add 50-100µl of elution buffer. Resuspend the beads by using pipette. Incubate in a metal bath at 65°C for 5-10 minutes.
13. Briefly spin down the tube. Place the tube on the magnetic rack for 5 minutes or until the beads have formed a tight pellet. Transfer the supernatant to a new centrifuge tube.
14. The solution can be stored at -20°C for a short period of time and at -80°C for a long time.